

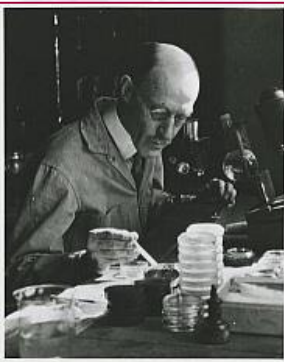


Techniques of cell and molecular biology

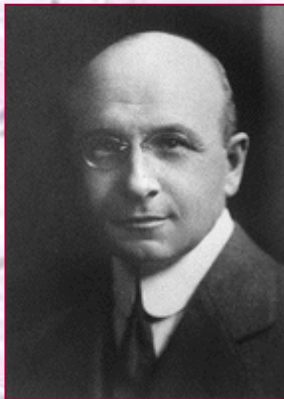
1. Cell fractionation
2. Autoradiography
3. X-ray crystallography
4. Histochemistry and cytochemistry
5. Immunohistochemistry
6. Hybridization techniques
7. Human brain connectivity



Vital observations



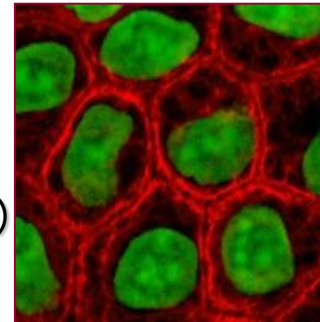
Ross Granville Harrison
(1870-1959)



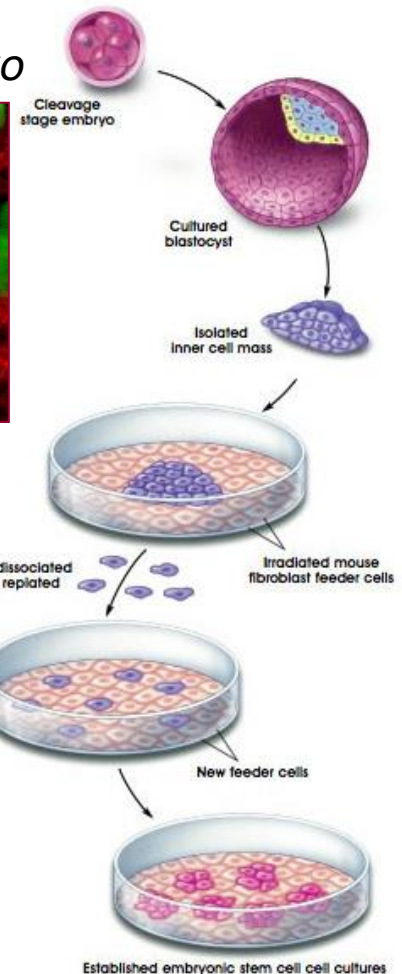
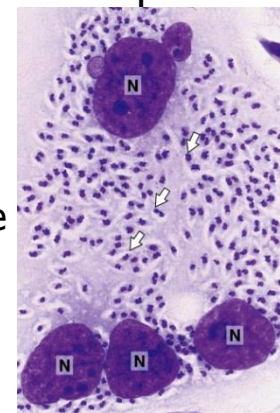
Alexis Carrel
(1873-1944)
The Nobel Prize
in Physiology or
Medicine 1912



- Cell, tissue and organ cultures: *in vitro* and *in vivo*
 - ✓ primary cell cultures:
 - dissociated (cell cultures)
 - explant (tissue cultures)
 - ✓ secondary: cell lines (HeLa cells)



- Medical applications:
 - ✓ Study of the metabolism of normal and cancerous cells
 - ✓ Development of new drugs
 - ✓ Study of parasites that grow only within cells, such as viruses, mycoplasma and some protozoa
 - ✓ Vaccine creation
 - ✓ Cytogenetic research:
 - chromosome analysis
 - determination of human karyotype
 - genetic disorders
 - gene and cell engineering



Cell fractionation



Albert Claude
(1899-1983)
Nobel Prize,
1974



Theodor Svedberg
(1884-1971)
Nobel Prize,
1926



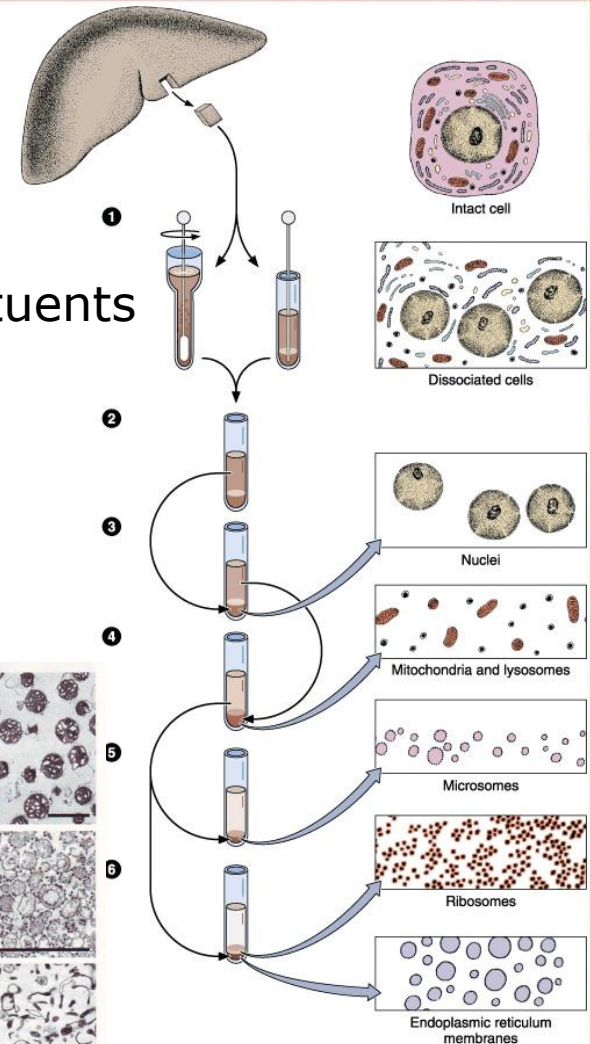
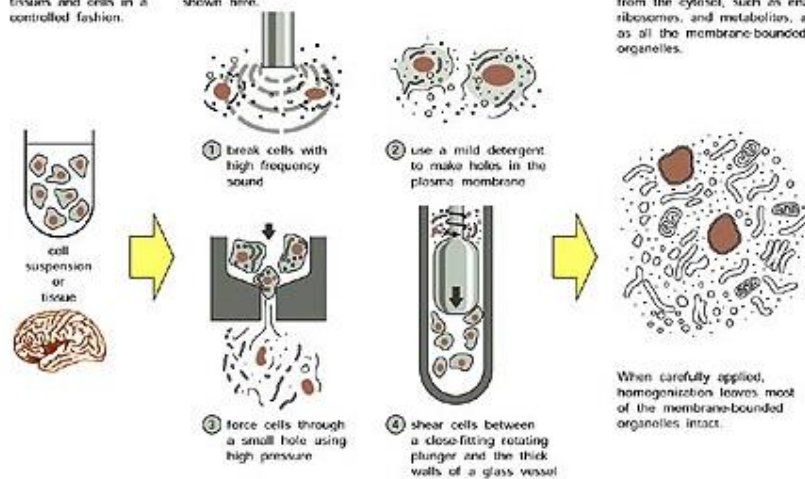
- ultracentrifuge – **T. Svedberg**
- cell fractionation – **A. Claude**
 - ✓ allows the isolation of cell constituents by differential centrifugation
- density gradient centrifugation

BREAKING CELLS AND TISSUES

The first step in the purification of most proteins is to disrupt tissues and cells in a controlled fashion.

Using gentle mechanical procedures, called homogenization, the plasma membranes of cells can be ruptured so that the cell contents are released. Four commonly used procedures are shown here.

The resulting thick soup (called a homogenate or an extract) contains large and small molecules from the cytosol, such as enzymes, ribosomes, and metabolites, as well as all the membrane-bounded organelles.



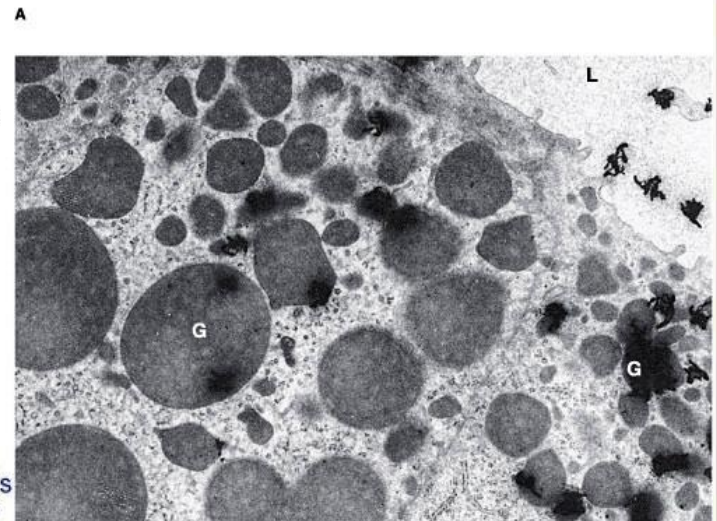
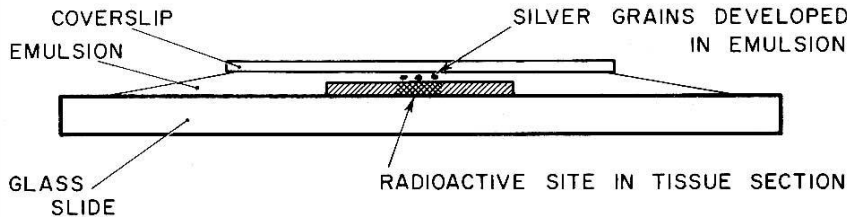
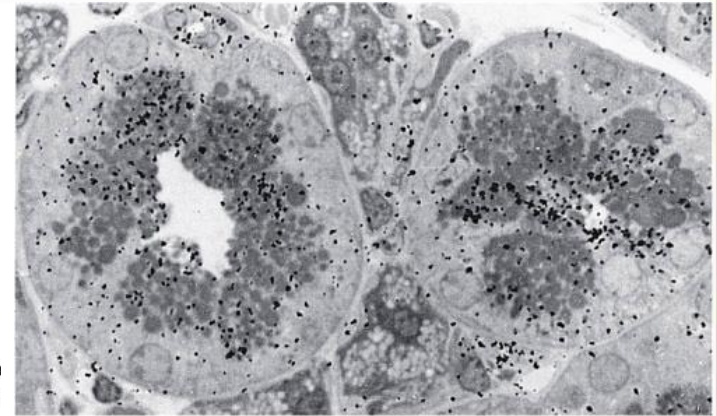
Autoradiography of tissue sections



Antoine Lacassagne
(1884-1971)

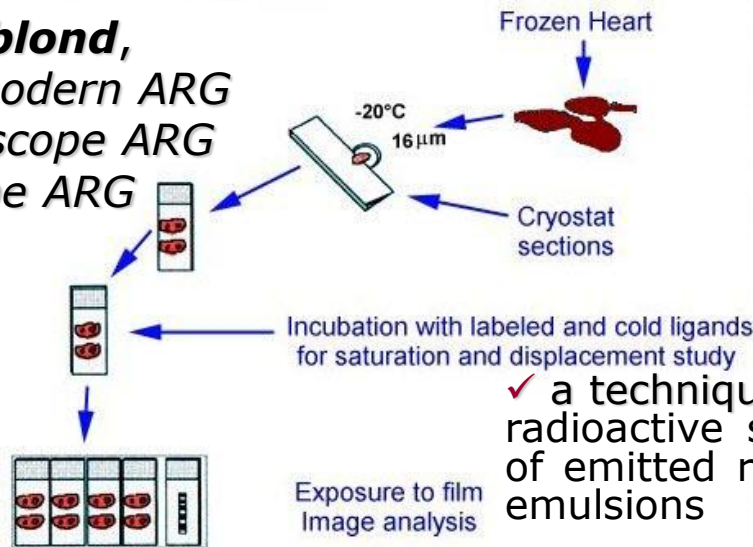
1924, developed the first autoradiographic method

Radioautography (Autoradiography)



Belanger and Leblond,
1946 – begin of modern ARG

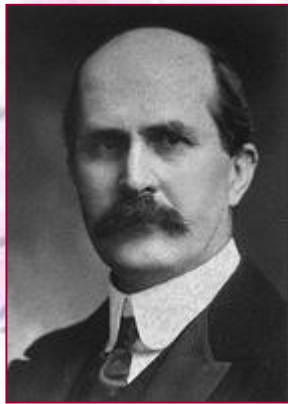
- ✓ electron microscope ARG
- ✓ light microscope ARG



✓ a technique that permits the localization of radioactive substances in tissues by means of emitted radiation effects on photographic emulsions



X-ray crystallography

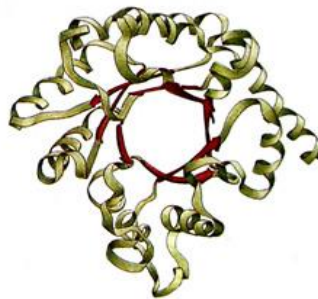
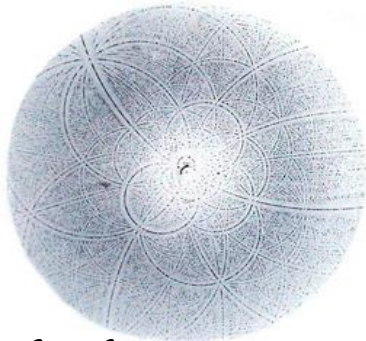


Sir William Henry Bragg
(1862-1942)



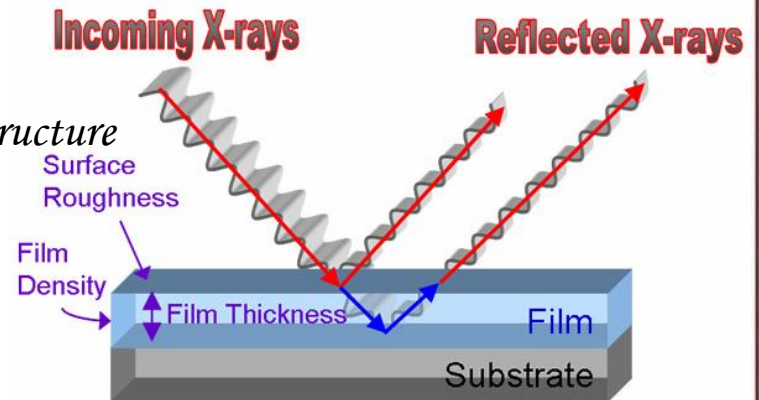
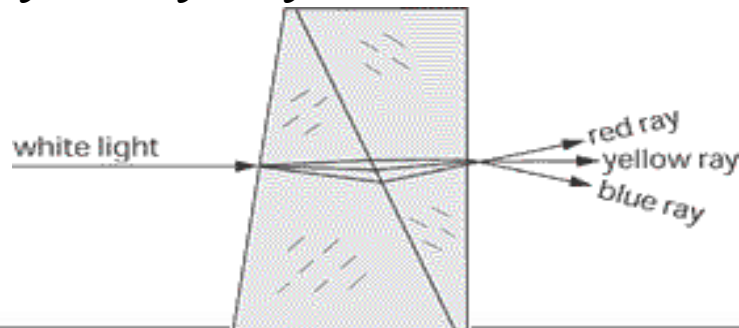
Sir William Lawrence Bragg
(1890-1971)
Nobel Prize in Physics, 1915

- ✓ a method of determining the arrangement of atoms within a crystal
- ✓ to solve the crystal structure of:
 - proteins
 - cholesterol and vitamin B12
 - hemoglobin and myoglobin etc.



W.H. Bragg and his X-ray spectrometer at University College, London.

"for their services in the analysis of crystal structure by means of X-rays"



Histochemistry and cytochemistry



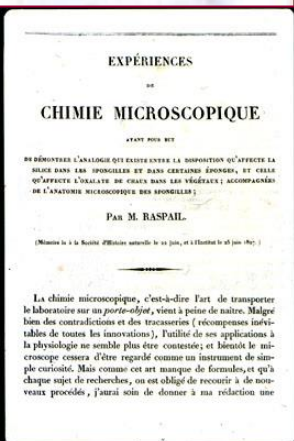
Founder of the method: *Francois-Vincent Raspail* (1794-1878)

- ✓ Histochemistry = LM results
- ✓ Cytochemistry = EM results

Quantitative analysis: principles

- ✓ **to preserve structure** of cells and tissues
- ✓ **localizations on the original sites in the cell:**
to avoid translocation
- ✓ **specificity of the reaction:**
positive and negative controls

Qualitative analysis: microspectrophotometry





Enzyme histochemistry: principles and applications

- Enzyme, substrate, product
- Principles:
 - ✓ fresh, unfixed material – cryostat
 - ✓ short-term fixation at lower temperature
 - ✓ pH optimum of the detected enzyme: buffers
- Basic requirements:
 - ✓ demonstration of final product, not the enzyme
 - ✓ insoluble product: true localization in the cell
 - ✓ color product: easily visible on the background



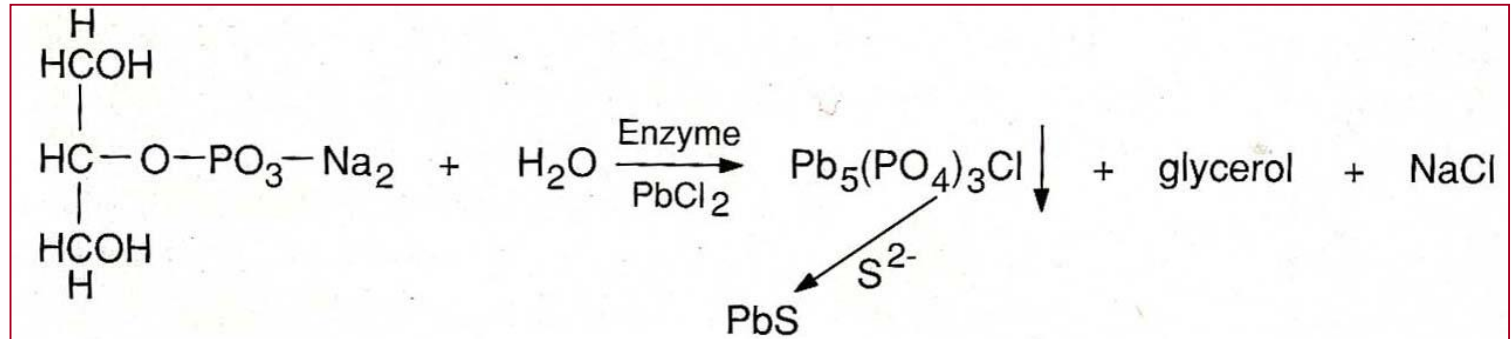
Enzyme + Substrate = unstained reaction product
Product + Dye = insoluble colored final reaction product



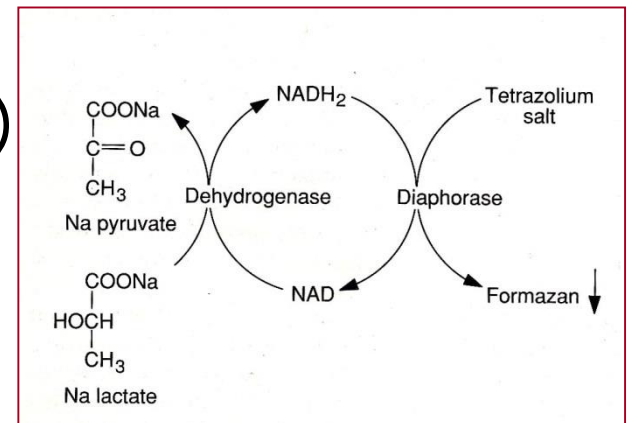


Catalytic enzyme histochemistry

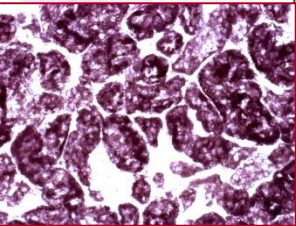
- Hydrolases: method of *Gomori* (1950, 1952)
 - ✓ acid phosphatase – lysosomes and Golgi complex



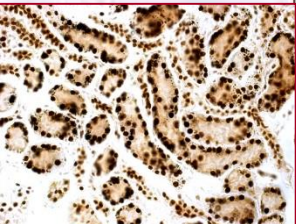
- oxidoreductases: method of *Nachlas* (1957)
 - ✓ succinate dehydrogenase – mitochondria
 - ✓ catalyzes the oxidation of succinate to fumarate



succinate + acceptor = fumarate + reduced acceptor



Rat kidney



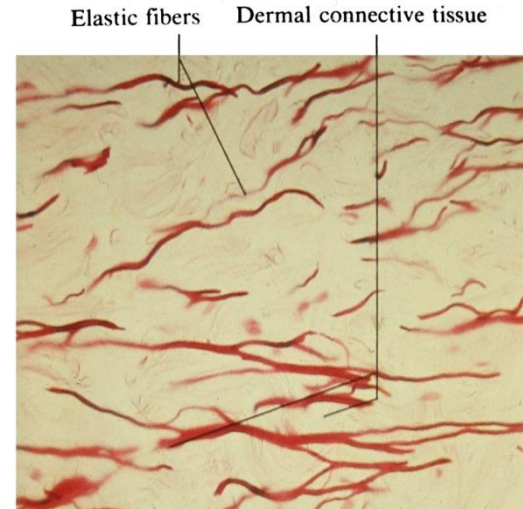
Rat kidney





Demonstration of proteins

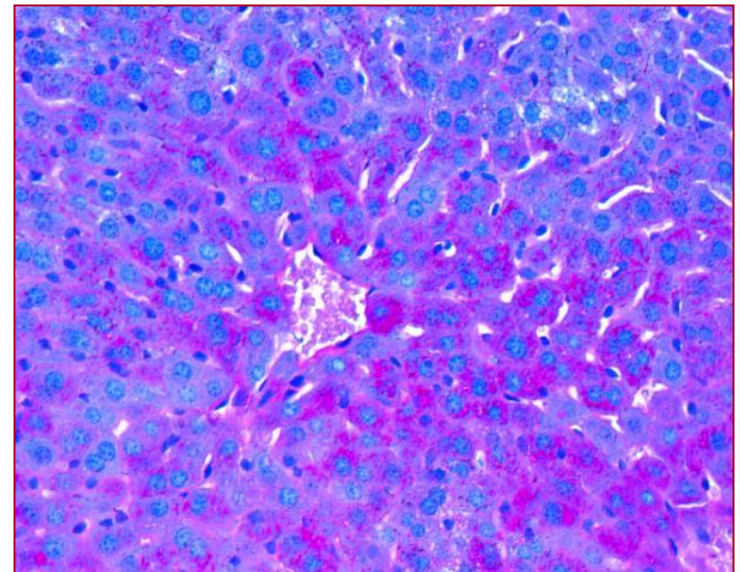
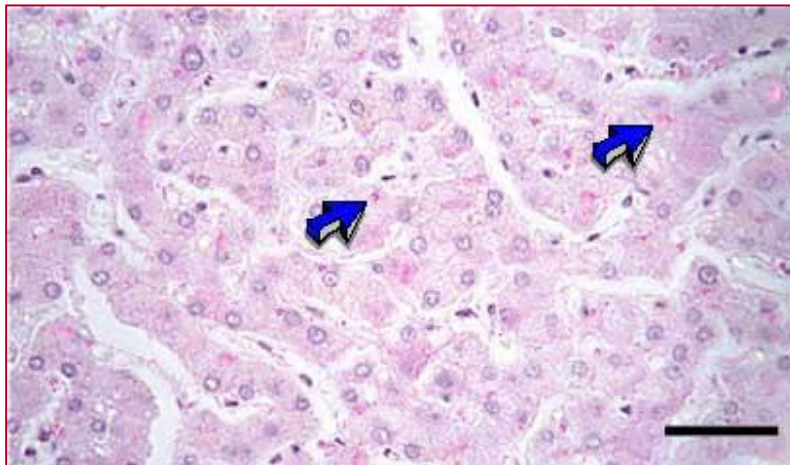
- the histochemical methods usually do not permit identification of specific proteins in cells and tissues
- elastic fibers:
 - ✓ orcein
 - ✓ *Weigert's* resorcin-fuchsin
- amino acids:
 - ✓ immunocytochemistry
- chemical groups:
 - ✓ paraldehyde-fuchsin – neurosecrete, insulin
- solubility and isoelectric point





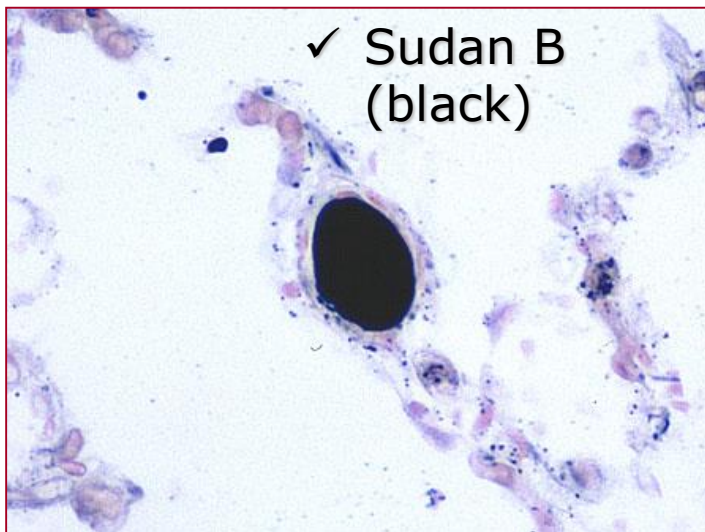
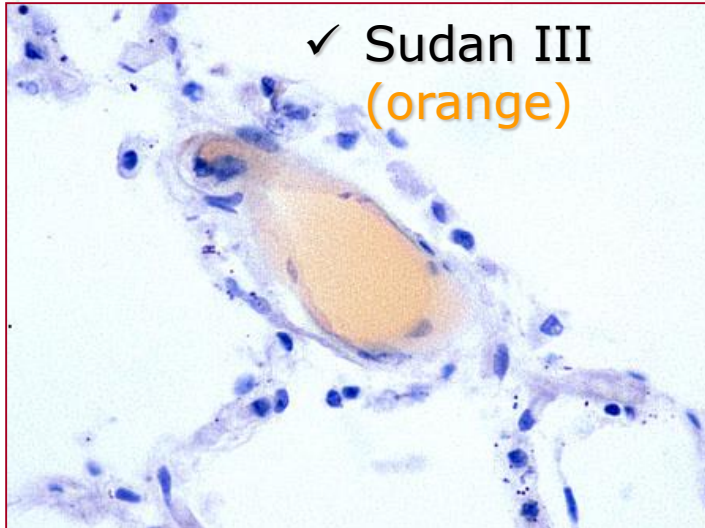
Demonstration of oligosaccharides & polysaccharides

- **PAS**-reaction (**P**eriodic **A**cid-**S**chiff)
 - ✓ demonstration of **glycogen** in tissues
 - ✓ demonstration of **glycoproteins**
 - ✓ demonstration of **glycosaminoglycans**

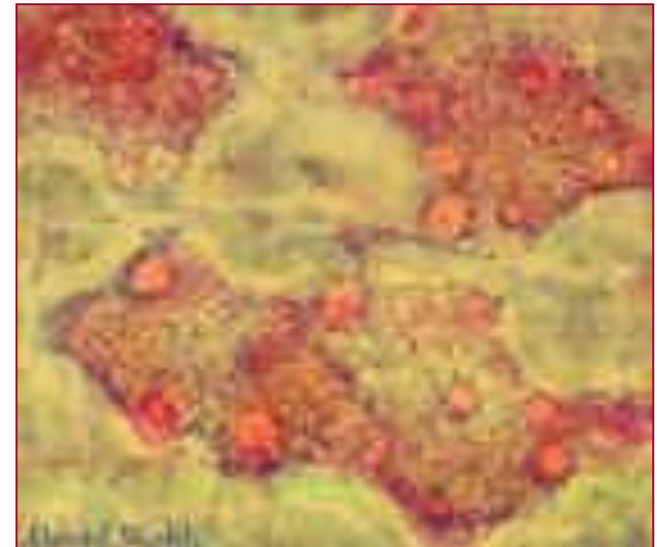




Demonstration of lipids



- best revealed with dyes that are soluble in lipids:



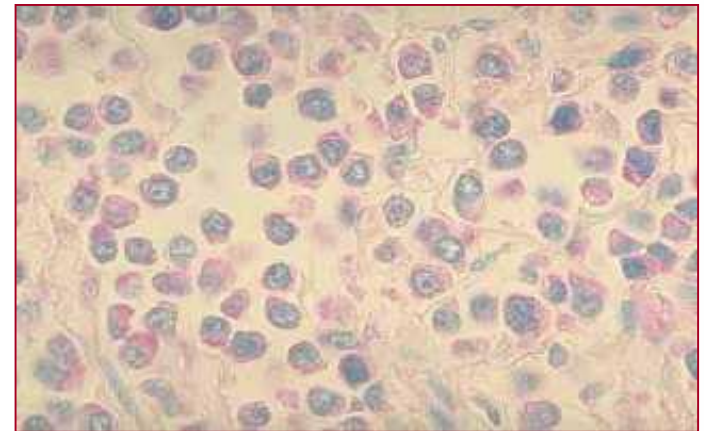
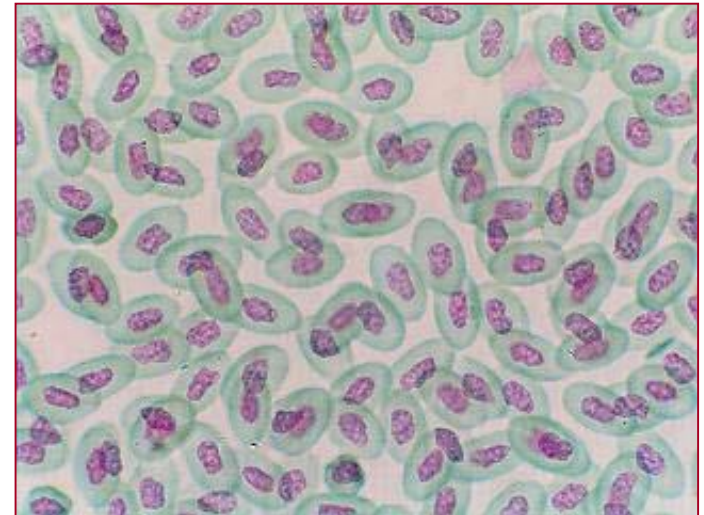
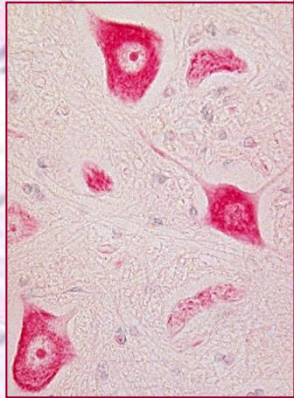
- ✓ Sudan IV
(red)





Demonstration of nucleic acids

- based on basophilia of nuclei acids
- ✓ *DNA*: method of *Feulgen* and *Rossenbeck* (1924) (*Feulgen* reaction)
- ✓ *RNA*: method of *Brachet* (1940-1941) methyl green-pyronin



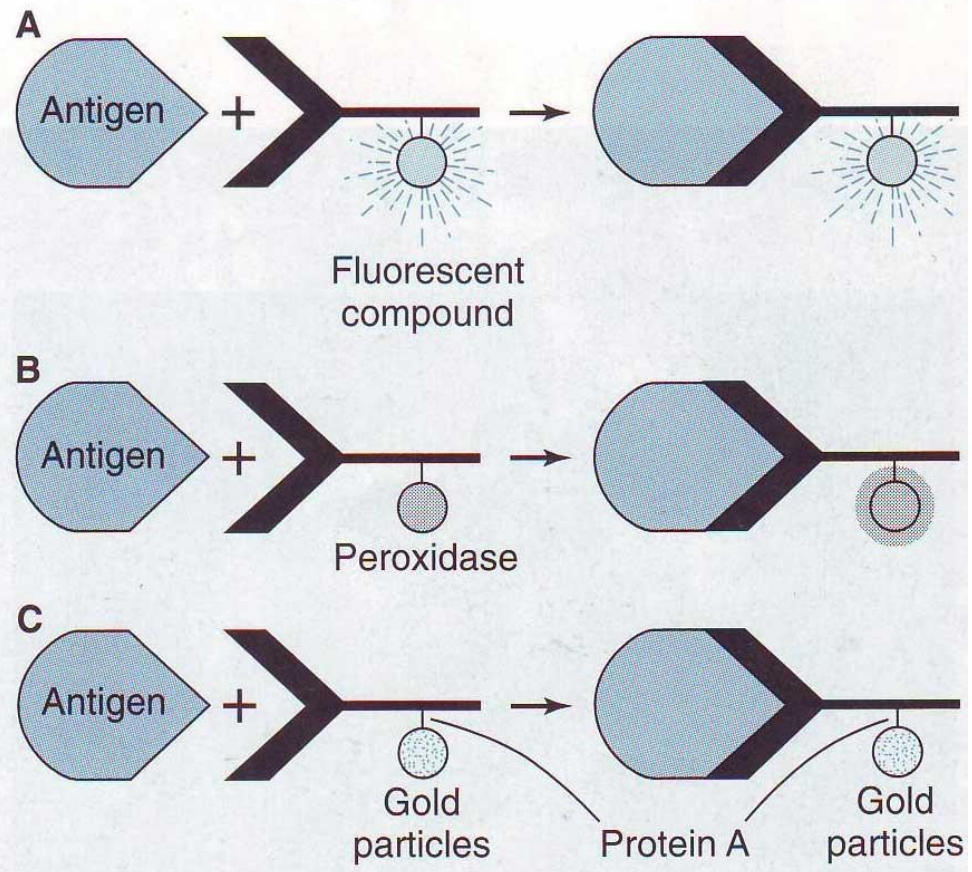
Immunohistochemistry

- based on an antigen-antibody reaction
- Methods of labeling antibodies:

✓ coupling with a fluorescent compound: immunofluorescence method – *Coons, 1941*

✓ coupling with an enzyme: PAP method (*Sternberger et al., 1970*) ABC technique (*Hsu, 1981*)

✓ coupling with gold particles



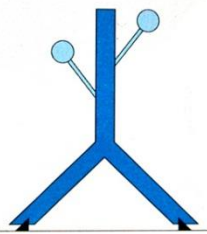
Immunohistochemistry

- based on an antigen-antibody reaction

- Methods of localizing of **antigens**:

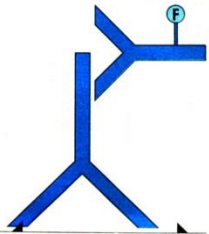
- ✓ direct method – detection with the fluorescence microscope
- ✓ indirect method – more sensitive but requires more steps

Direct immunoenzyme-conjugate method

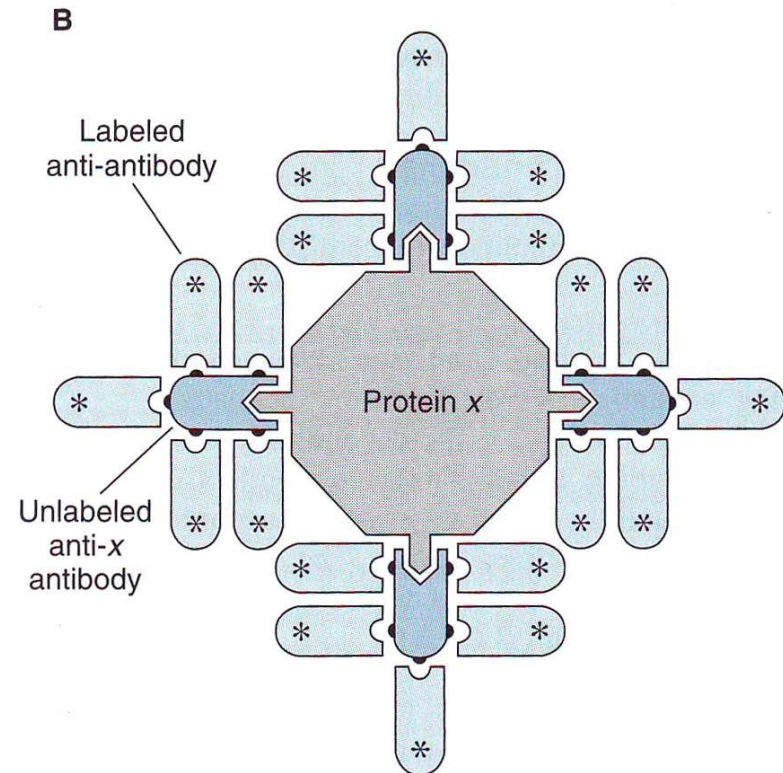
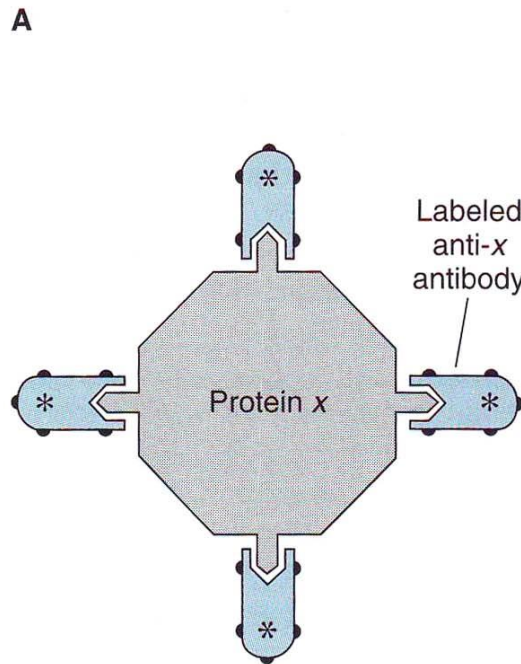


Enzyme
Primary Antibody
Antigen

The indirect immunofluorescence method

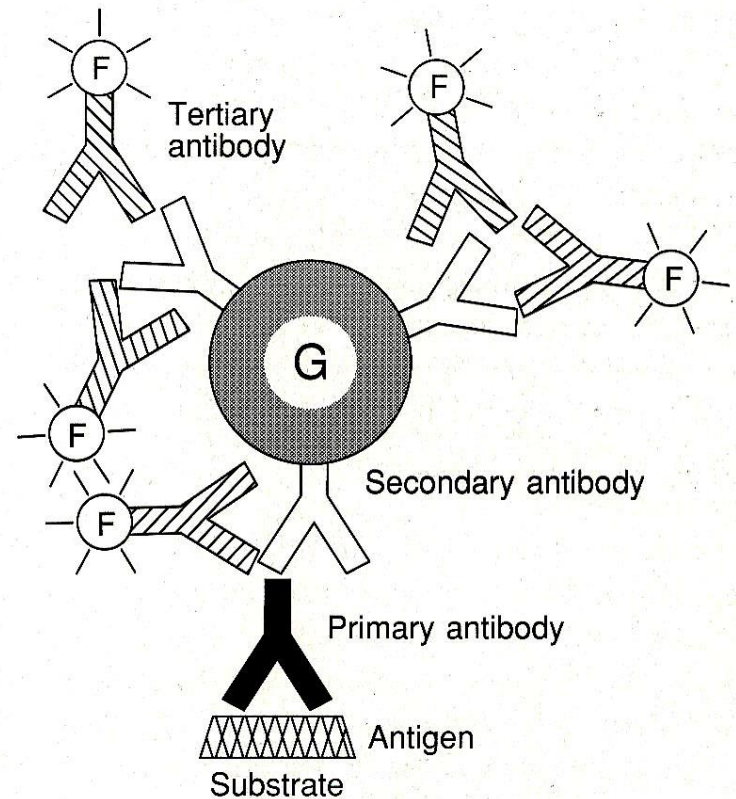
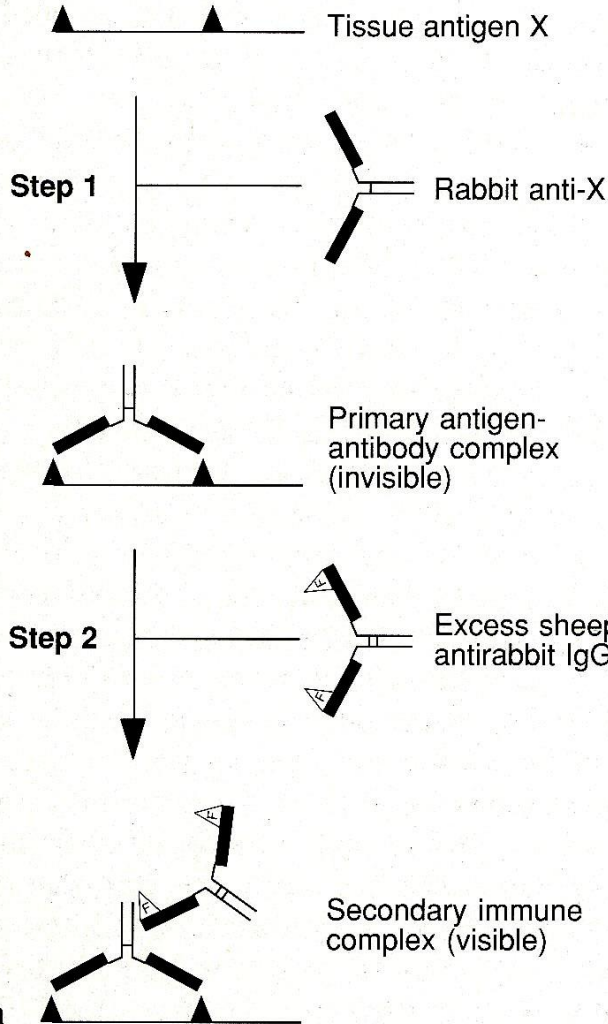
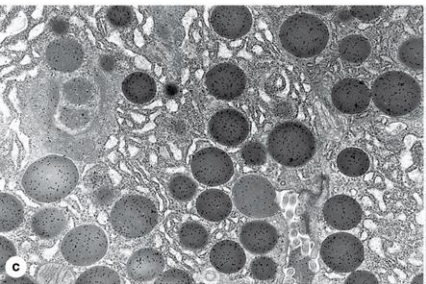
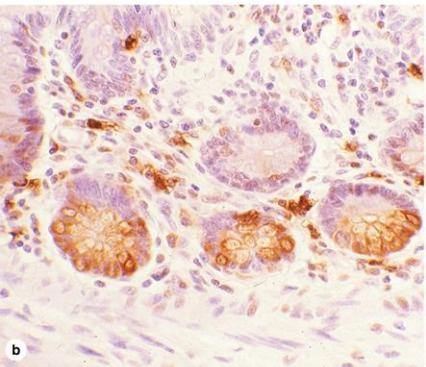
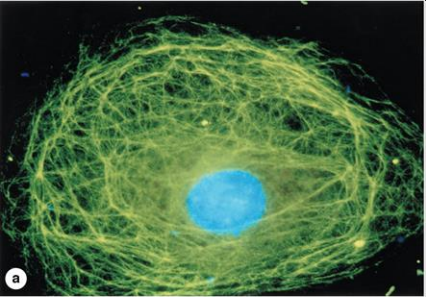


Indirect fluorescence method (F)



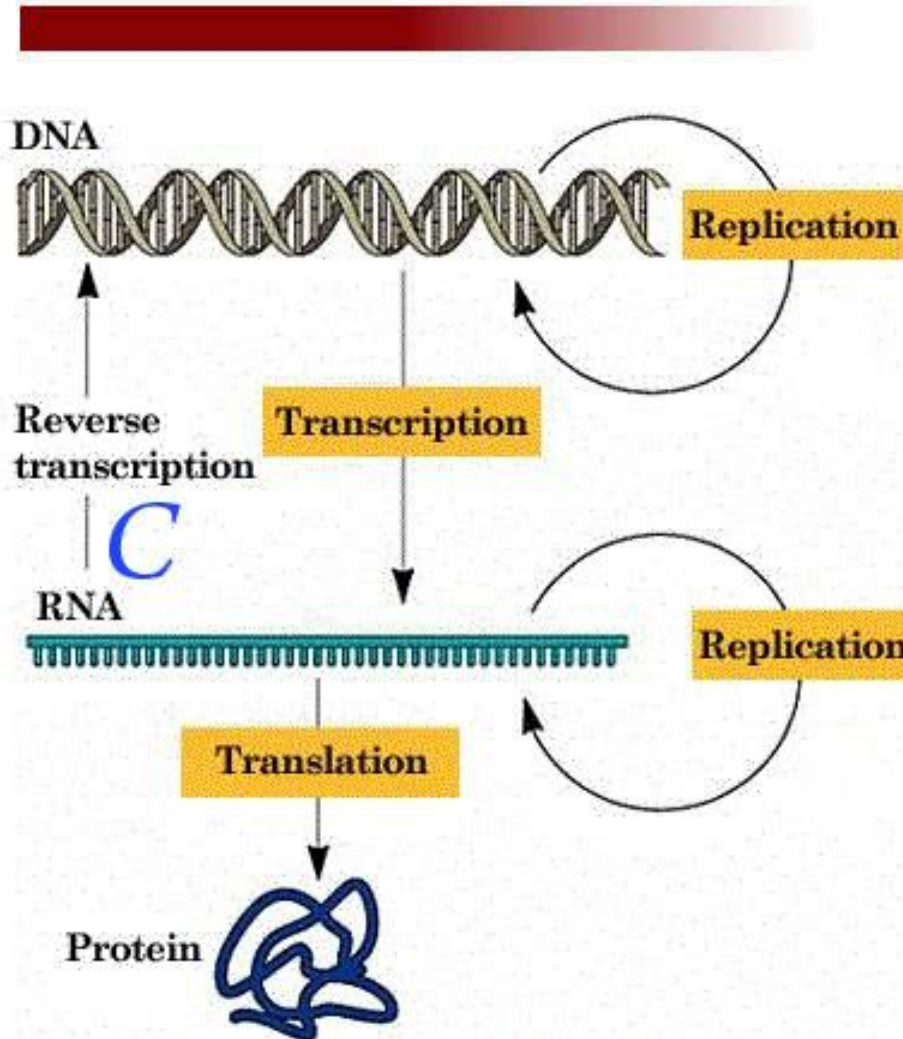


Immunohistochemical technique





Hybridization techniques



Southern blotting – detection of a specific DNA sequence, *Edwin M. Southern*

Northern blotting – detection of RNA fragments (or isolated mRNA)

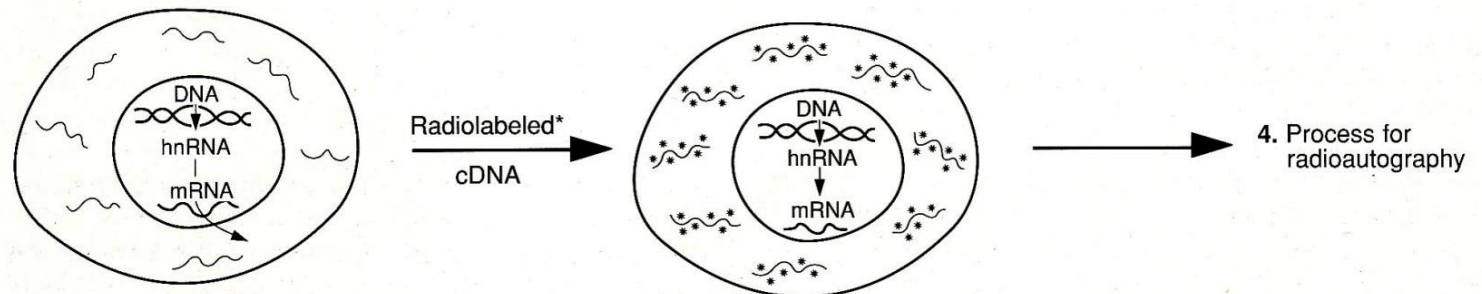
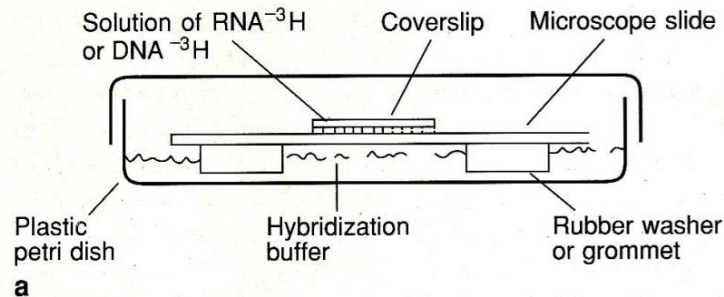
Western blotting – detection of specific proteins





In situ hybridization

■ Radioactive *in situ* hybridization (ISH):



1. Tissue sections or cells cultured on slides
2. Fixation

3. Hybridize radiolabeled cDNA probe with complementary mRNA molecules
4. Process for radioautography

b



In situ hybridization

- Nonradioactive *in situ* hybridization:

✓ originally developed by *Pardue and Gall* (1969), and (independently) by *John et al.* (1969)

Preparation of slides and fixation of material

Choice of the probe and its labeling

**Denaturation of in situ target DNA
(probe and target)**

***In situ* hybridization**

Immunocytochemical visualization

Microscopy

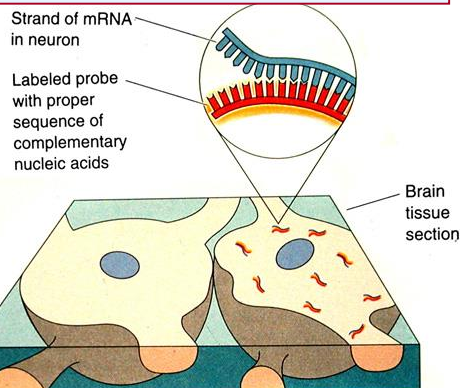
Flow diagram for ISH procedure

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Prof. Dr. Nikolai Lazarov



Nonradioactive *In Situ* Hybridization

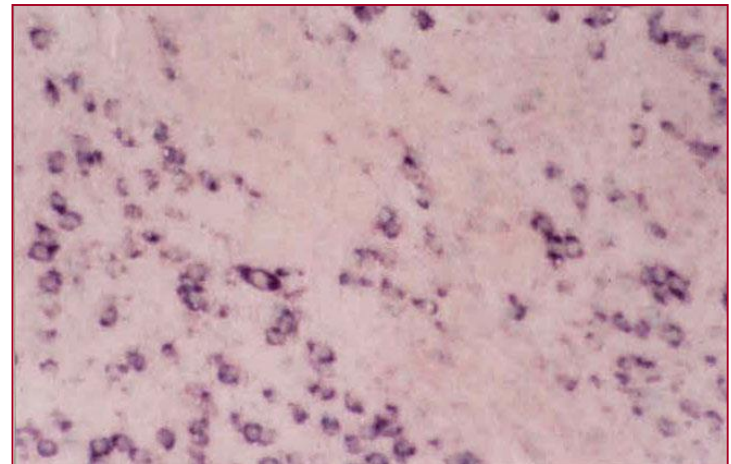
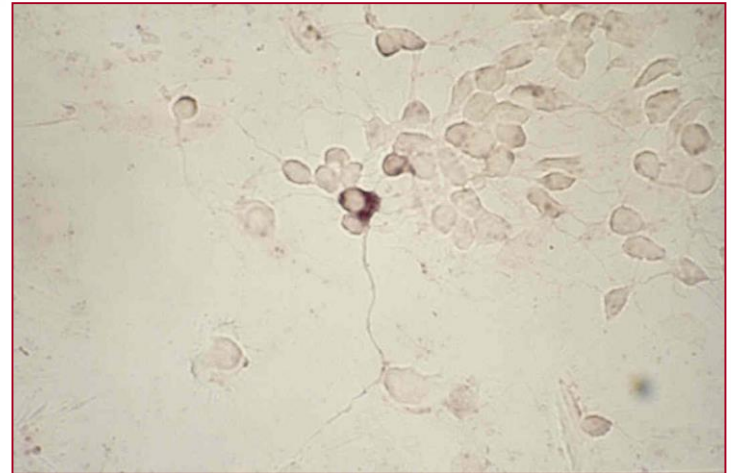




Medical applications

- In **fundamental research**:
 - ✓ gene mapping
 - ✓ localization of gene expression
 - ✓ systematization of nuclear DNA and RNA
 - ✓ replication
 - ✓ cell sorting

- In **clinical research**:
 - ✓ cytogenetics
 - ✓ prenatal diagnostics
 - ✓ gene disorders
 - ✓ diagnostics of infectious and malignant diseases
 - ✓ biological dosimetry



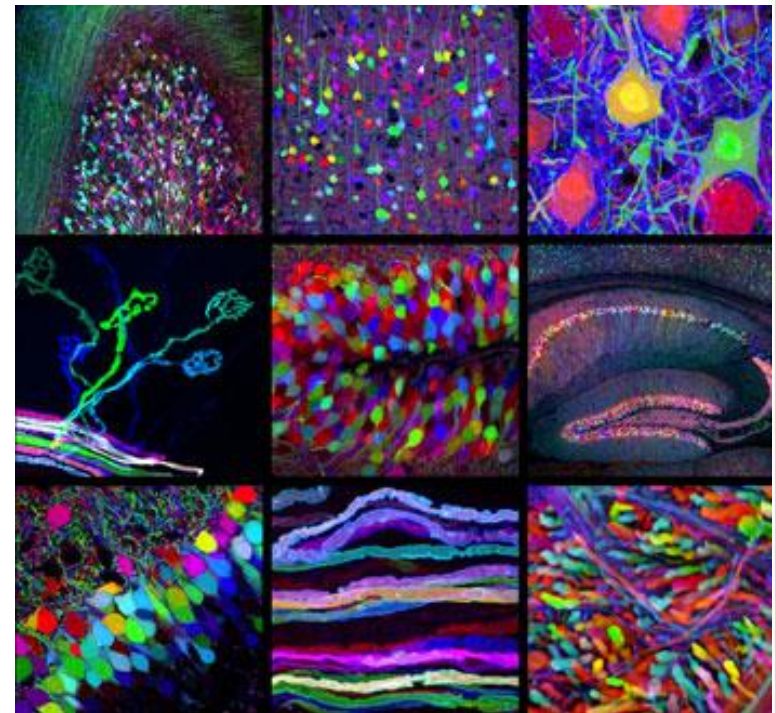
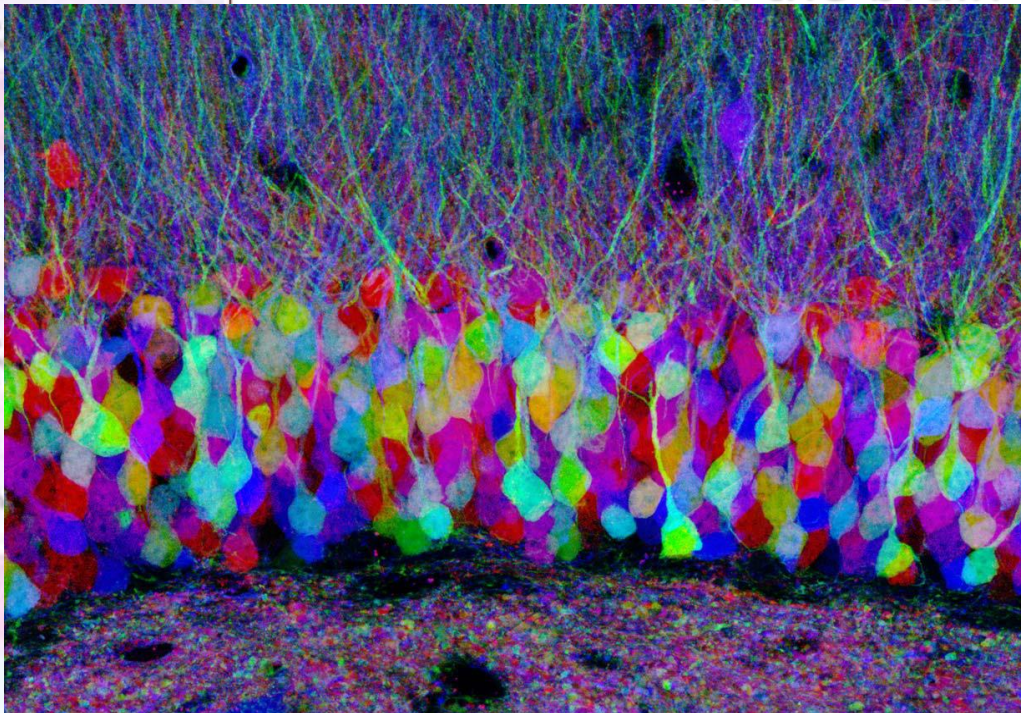
Human brain connectivity

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LICHTMAN
LAB



- Connectome
- Brainbow
- ✓ transgenic painting in the brain



Lichtman et al.: *Nature* 2007, 450:56-62

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Prof. Dr. Nikolai Lazarov



Thank you...

medicine

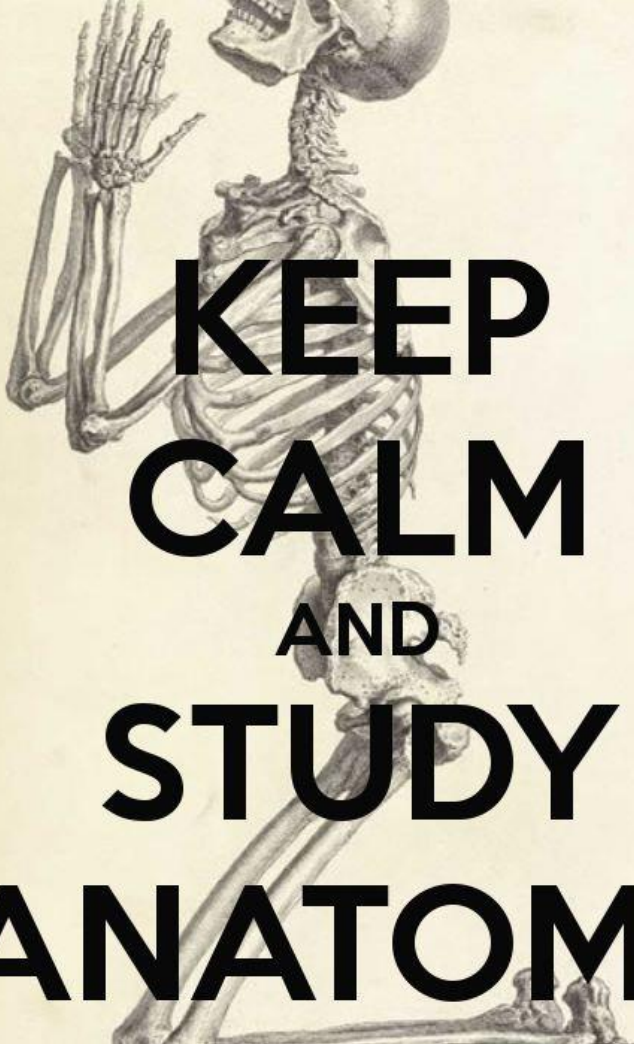
Faculty of medicine

EXIT

ENTRY



No Pain, No Gain



**KEEP
CALM
AND
STUDY
ANATOMY**

